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A Simple Combination of Active and Smart Packaging in Extending and Monitoring the Meat Quality Stored at Cold Temperatures ($4 \pm 1^\circ\text{C}$)

Abstract

A simple combination of active and smart packaging has been developed using garlic extract as the raw material as well as bromothymol blue (BTB) and phenol red (PR) as the solution. Therefore, this study aims to extend and monitor the beef quality deterioration at temperatures of $4 \pm 1^\circ\text{C}$ by applying a double function in active and smart packaging. The active packaging uses garlic extract with concentrations of 0%, 15%, and 20% (w/w) to release anti-microbial agents. Meanwhile, the smart paper uses a combination of BTB and PR solutions at pH 5.00. On the ninth day, beef packed without adding 15% and 20% garlic extract on active paper is unfit for eating. The smart indication label's color change profile is as follows: dark yellow (fresh), reddish-yellow (to be consumed immediately), and faded red (rotten). The color change of the smart BTB+PR pH 5.00 indicator label in response to all meat deterioration criteria demonstrates a linear correlation for determining the extent of rottenness during storage. Therefore, active paper and smart indicator labels are combined to extend the shelf life and monitor meat quality deterioration.

1. Introduction

Meat is one of the most basic nutrients containing high levels of easily digestible protein, calorie-dense fat, vitamins, and other micronutrients. Its high nutritional value corresponds to the rising demand as the world's most significant food product during the past several years.¹ The high demand should be accompanied by an increase in the fresh meat grade, including the quality and safety to minimize damage due to chemical, physical, and microbiological factors. The decrease in quality is due to the high fat and water content, making it susceptible to microbial contamination and lipid oxidation. Furthermore, quality degradation occurs at several stages of the production chain, including preparation, storage, and distribution.^{2,3} The use of active packaging in conjunction with cold storage is one of the efforts that can be made to prolong the shelf life and enhance the quality.

Active packaging applications can be used to inhibit microbial growth in meat.⁴⁻⁶ These applications incorporate specific active substances into packaging methods to preserve food products from bacteria that cause spoilage during storage.⁷ The spoilage inhibition mechanism can be conducted in two ways of direct and indirect contact. The direct contact is between packaging materials and food, while in the indirect contact, the active packaging releases volatile anti-microbial agent.^{8,9} Volatile anti-microbial compounds are essential oils derived from herbs and spices, and the anti-microbial activities of different species of *Allium* have been investigated.¹⁰⁻¹² Furthermore, Zheng Dong et al. (2019) developed an active packaging material based on a PP/LDPE composite film containing *Allium sativum* essential oil. The addition of essential oils into the packaging extends the shelf life of large yellow croaker (*Pseudosciaena crocea*) fillets for five days stored at $4 \pm 1^\circ\text{C}$.¹³ Seydim and Sarikus (2006) also reported that the anti-microbial properties of whey protein isolate containing essential garlic oil with a ratio of 3.0 – 4.0% (wt/vol) effectively inhibited the growth of *Salmonella Enteritidis*, *S. aureus*, *E. coli* O157:H7, *L. monocytogenes*, and *Lactobacillus Plantarum*.¹⁴ The use of garlic extract in active packaging can inhibit microbial growth due to its volatile *Allicin* constituent.¹⁵ Allicin chemicals suppress microbial development by increasing their permeability in penetrating bacterial cell walls to decrease protease enzyme production and disrupt protein and nucleic acid metabolism.¹¹

The application of active packaging on fresh meat can be accompanied by the addition of smart packaging in the form of indicators that monitor changes in quality. This is achieved through chemical reactions between indicators and the results of microbial metabolism or changes in the meat's chemical composition. Smart packaging incorporates indicators that convey information about the product's quality without causing damage to the packaging. This reduces the risk of loss due to product damage and provides a more accurate condition estimate than conventional expired labels. Furthermore, smart packaging also assists distributors in adjusting the prices of the products sold.¹⁶ For example, lowering prices when quality has decreased to avoid losing food products. Previous studies applied this method to avocados (Dirpan et al., 2021)¹⁷, tuna fillets (Agustianti et al., 2021)¹⁸, shrimp (Wu et al., 2021)¹⁹, cherry tomatoes (Kim et al., 2021)²⁰ and *arummanis* mango (Dirpan et al., 2018)²¹. According to Shukla et al. (2015), color-based pH indicators can be used as smart packaging to monitor the presence of volatile amines from microbial metabolites in meat spoilage. The increase in pH during decay causes a significant change in the indicator's color to be visually observed.²² Meanwhile, Romero et al. (2019) evaluated the quality of packaged cow's milk using two smart packaging prototypes based on the bromothymol blue pH indicator.²³ Julyaningsih et al. (2020) and Yolanda et al. (2020) combined active and smart packaging based on pH indicators methyl red and bromothymol blue to monitor the tuna fillet's quality.^{24,25} Meanwhile, Yong and Liu (2020) combined anthocyanin-rich extracts into biopolymer-based films to extend shelf life and monitor the food product's quality.²⁶

The spoilage and contamination of food supplies jeopardize global food security. Therefore, the innovative application of active and smart packaging needs to be developed. As a result of these considerations, the purpose of this research is to evaluate the efficacy of active paper with garlic extract addition when applied to fresh beef during storage at low temperatures ($4\pm 1^{\circ}\text{C}$). Furthermore, it investigates the relationship (correlation) between color analysis of smart packaging indicators on various parameters of the beef spoilage test.

2. Material and Methods

2.1. Materials

The beef tenderloin is purchased fresh at the slaughterhouse, Antang. Meanwhile, garlic as an anti-microbial compound in the activated paper was purchased from supermarkets. Bromothymol blue (BTB) and phenol red (PR) color indicators in smart packaging were purchased from Sigma-Aldrich (Merck) with Mueller Hinton Agar (MHA) and Nutrient Agar (NA) media. This research also uses *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*, and the other chemicals used were of analytical grade.

2.2. Garlic Extract Preparation

Garlic of 50 g was blended until smooth and put into a container containing 250 ml of 96% alcohol with 3 x 24-hour immersion. The stirring process was conducted 14 times in 5 hours during immersion. Subsequently, it is separated from the garlic dregs and the filtrate by filtering. At 45°C , the filtrate was evaporated with an industrial rotary evaporator (TMAX-2L-1) until it became thick reddish-yellow garlic.²⁷

2.3. Active Paper Preparation

Active paper preparation was carried out using Whatman paper no. 1 cut into small pieces of 15 g and soaked in 250 ml of distilled water for 24 hours. The paper soak was added with 250 mL of distilled water and crushed using a blender for 20 minutes until a pulp was formed. The paper pulp obtained was squeezed to remove the water, and 30% (w/w) tapioca starch was dissolved in 50 mL of distilled water, then homogenized with the pulp. Furthermore, 100 mL of acetic acid (1%) was added with 0.45% chitosan powder and homogenized with a blender for 5 minutes. Garlic extract with concentrations of 15% and 20% (w/w) on filter paper was added with 50 mL of distilled water, respectively. Furthermore, 0.205 g of tween 80 was added and mixed at room temperature using a hotplate magnetic stirrer to generate an emulsion. The paper dough is poured over the surface of the printer container following the styrofoam size used as a beef storage container and flattened to make a wet paper sheet. Meanwhile, the wet paper sheet on the filter surface is pressed between the glass surfaces with a load of 2 kg. It was then dried at 40°C for 48 hours, and a paper turning procedure was after 24 hours of drying (Modification of Wiastuti et al., 2016).²⁸

2.4. Indicator Solution Preparation

BTB and PR (1:1) indicator solutions were made with 1% (w/v) and dissolved in 95% ethanol. Furthermore, the indicator was adjusted to pH 5 using a solution of glacial acetic acid (Kuswandi et al., 2017).²⁹

2.5. Smart Indicator Label Preparation

Whatman paper no. 1 was cut with a size of 2 x 4 cm and immersed in 20 ml of indicator solution for 12 hours at room temperature (28±2°C). The indicator labels were dried using an electric dryer and a closed container with a drying distance of 30 cm.

2.6. Application of Active and Smart Packaging on Fresh Beef

Fresh beef (tenderloin) with normal pH (5.6 – 5.7) was obtained from the Tamangapa Antang Makassar abattoir, which was taken at a relative postmortem time of about 3 hours. The meat was packaged in plastic (PP) containers and put in a cooler, then immediately transported to the laboratory and prepared sterilely into 180-gram pieces/packages. Activated paper with different concentrations was inserted into the styrofoam base. Furthermore, the study used A0: Control (without garlic extract addition); A1: 15% garlic extract; A2: 20% garlic extract. The pieces of meat were packed in styrofoam (1.05 g/cm³) covered with active paper that filled the entire base. The indicator label was placed inside the package by sticking to the LDPE film surface (0.9 g/cm³) used as a cover. Observation of beef (tenderloin) packaged with smart and active packaging was conducted at a cold temperature (chiller) of 4 ± 1°C every 3 x 24 hours for 21 days of storage. The design for implementing smart and active packaging can be seen in Figure 1.

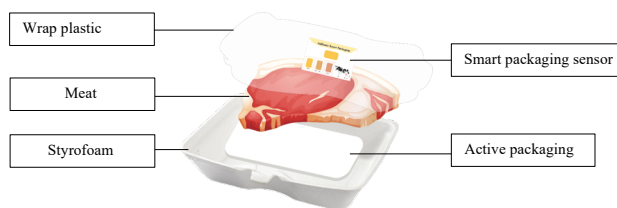


Figure 1. Active and Smart Packaging Design

2.7. Antibacterial Activity Testing of Agar Diffusion Method on Activated Paper

The antibacterial activity of active paper was tested to determine the inhibition toward the growth of gram-positive bacteria *Staphylococcus aureus* (ATCC 29213), gram-negative bacteria *Escherichia coli*, and *Pseudomonas aeruginosa*. Sheets of active paper with a diameter of 5.5 mm were placed on Mueller Hinton Agar (MHA) media, which spread 0.1 ml of microorganism culture on its surface. Furthermore, the Petri dishes were incubated at 37°C for 24 hours, and after incubation, a clear zone (inhibition zone) was formed around the active paper. The resistance diameter was measured in millimeters (mm) using a caliper³⁰, and each test was conducted in 3 replications to obtain the average result.

2.8. Parameters of Beef Observation packaged with a combination of active and smart packaging

2.8.1. pH Measurement

pH measurements measured the initial and final pH during storage of fresh beef using Horiba Laquatwin Compact pH Meter P-33 with an accuracy of 0.01%. A total of 1 g of the sample was mashed and dissolved in 10 ml of Aquades. Furthermore, it was inserted on the surface of the pH meter sensor until the value was printed on the pH meter screen.

2.8.2. TVBN Value Analysis

The meat sample was weighed at $1 \text{ g} \pm 0.1 \text{ g}$, pulverized in a mortar with 3 ml of 7% TCA solution, and filtered to obtain the filtrate. The boric acid solution of 1 ml was put into the "inner chamber" of the Conway cup, and the lid was placed in a position to cover the cup. Furthermore, the filtrate was fed into the outer chamber on the left. Then 1 ml saturated K₂CO₃ solution was added to the outer chamber on the right, ensuring that the filtrate and K₂CO₃ do not mix. The cup was instantly closed and rotated to mix the two liquids in the outer chamber. The blank solution was prepared the same way as the standard solution, except that the filtrate was replaced with a 7% TCA solution. Subsequently, it was stored at $28 \pm 2^\circ\text{C}$ for 24 hours, and the boric acid solution in the inner chamber containing the blank was added with two drops of color indicator solution, then titrated with 0.02 N HCl until it turned pink. Conway cup containing the filtrate was added with two drops of color indicator solution and titrated with the same solution until it turned pink such a blank (SNI-2354.8:2009; AOAC, 1995). The formula for determining TVBN is:

$$TVBN \text{ level (mg/100g)} = \frac{(V_c - V_b) \times N \times 14.007 \times 100}{w}$$

Description:

V_c = volume of HCl solution in sample titration

V_b = volume of HCl solution in blank titration

N = normality of HCl solution

W = sample weight (g)
14.07 = atomic weight of nitrogen

2.8.3. Calculation of Total Bacterial Count

Total bacteria were calculated based on the cup count method (SNI 2897:2008). The sample solution was made from 1 g of meat homogenized with 9 ml of sterile physiological solution (0.85% NaCl). A total of 1 ml of the sample solution was put into the first test tube containing 9 ml of the physiological solution until homogeneous, referred to as the 10^{-1} dilution. Then, the dilution was conducted up to a concentration of 10^{-6} and 1 ml from dilutions 10^{-4} , 10^{-5} , and 10^{-6} were put into separate sterile Petri dishes in duplicate. Similarly, \pm 15 ml of sterile NA media was homogenized and incubated at 30°C for 48 hours. The calculation of the Total Plate Count is:

$$N = \frac{\Sigma C}{[(1 \times n_1) + (0.1 \times n_2) + \dots] \times (D)}$$

Description:

N = number of colonies per ml/ per gram of product
 ΣC = total number of colonies counted
 n_1 = number of cups in the first dilution
 n_2 = number of cups in the second dilution
D = first dilution calculated

2.9. Smart Indicator Color Measurement Quantification

The color measurement of the smart indicator label was analyzed using a Digital color meter (T-135). This quantification is conducted by attaching the colorimeter sensor to the smart indicator label. The tool will then show the values of L (lightness), a (redness), and b (yellowness) on display. These three values are international standards of color measurement published by the Hunterlab Association Laboratory (2008). Furthermore, the smart packaging indicator color is determined by calculating the $^{\circ}$ Hue value using the formula³¹:

$$^{\circ}\text{Hue} = \tan^{-1} \frac{b}{a}$$

Description:

$^{\circ}\text{Hue}$ = parameters for color range
a = is a red-green mixed color
b = is a red-green mixed color yellow-blue

2.10. Level of Relationship (Correlation) between Test Parameters

The smart packaging indicator quantification data will be compared with the data for each beef spoilage parameter presented in a graph using the Sigma Plot software version 14.0.

2.11. Statistical analysis

All data from the test parameters were analyzed using analysis of variance (ANOVA) with Duncan's multiple range test using SPSS software version 22.0 (IBM Corp., United States), and the calculated values differed significantly when $p < 0.05$. In addition, the relationship

(correlation) of intelligent indicator color parameters to all beef spoilage parameters was presented in one graph using Sigma Plot software version 14.0.

3. Results and Discussion

3.1. The antibacterial analysis results of active paper

Antibacterial activity on active paper was tested against several bacteria that generally cause damage to beef, namely *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. Figure 2 illustrates the active paper's analytical findings for the bacterial inhibition zone region.

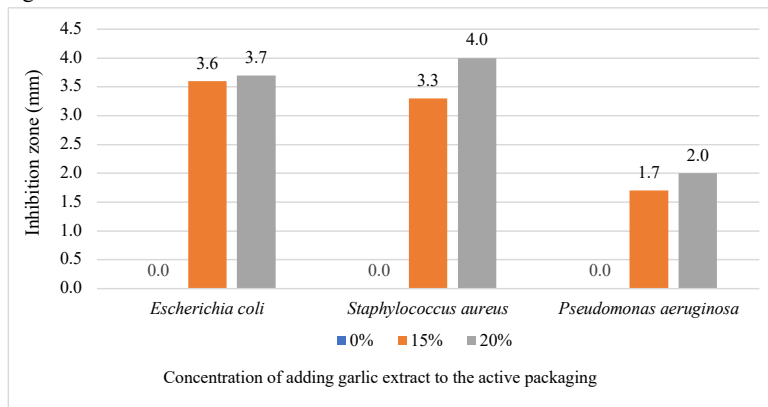


Figure 2. Diameter of Bacterial Growth Barriers on Activated Paper

The garlic extract addition on active paper had a significant effect ($p < 0.05$) on the growth inhibition of *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* bacteria (Figure 2). On the other hand, adding active paper without the garlic extract (0%) did not show any inhibition zones. In comparison, the addition of 15% and 20% extract significantly affected bacterial inhibitory activity, therefore, the garlic extracts have antibacterial activity.

The results obtained for activated paper with the addition of 15% and 20% garlic extract against *E. coli* and *S. aureus* bacteria were classified as a moderate inhibitory response to bacterial growth. Meanwhile, *P. aeruginosa* was classified as a weak inhibitory response to bacterial growth. This is consistent with Pan et al. (2009)'s assessment of the response to bacterial growth inhibition. The antibacterial activity with an inhibitory zone > 6.00 mm, 3-5 mm, and < 3 mm was considered strong, moderate, and weak.³²

The difference in the diameter of the inhibition zone formed on *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* was influenced by differences in the structure of bacteria that affect the penetration of cell walls by Allicin compounds. *E. coli* and *P. aeruginosa* are gram-negative bacteria with complex cell walls, therefore, the antibacterial compounds in garlic extract cannot inhibit bacterial growth properly. Gram-negative bacteria have three-layered cell walls consisting of lipoproteins, outer membrane phospholipids, lipopolysaccharides, and the lipid content of the cell walls, which ranges from 11-22%. The

phospholipid outer membrane makes it difficult for antibacterial chemical components to penetrate the cell walls of gram-negative bacteria. Meanwhile, *S. aureus* is a gram-positive bacterium with single or simple layered cell walls and a 1-4% lipid content. The low lipid content in the cell membrane causes antibacterial compounds to penetrate the bacterial cell wall. Therefore, their inhibitory power against bacteria is better than gram-negative bacteria. This is consistent with Vijayakumar et al.'s (2019) statement that differences influence the bacteria susceptibility to garlic components in the bacteria structure.³³ *Escherichia coli* bacteria cell membrane consists of 20% lipid, while *Staphylococcus aureus* consists of only 2% lipid. Differences in the lipid content of the bacterial membrane can affect the permeability of allicin and other elements. Salima (2015) confirms that garlic's antibacterial properties are greater against gram-positive bacteria (*Staphylococcus aureus*) than against gram-negative (*Escherichia coli*).³⁴

3.2. Meat pH

Analysis of the meat pH value determines the effect of adding various concentrations of garlic extract during cold storage. Therefore, the degree of acidity (pH) is an indicator to examine the meat freshness level. Figure 3 shows the pH changes that occur in beef during storage.

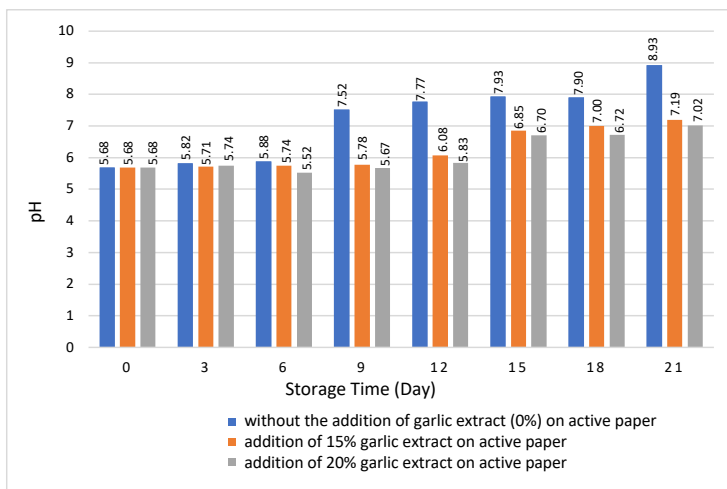


Figure 3. pH value of meat during cold storage

Figure 3 shows that the garlic extract addition with different concentrations affects increasing the pH value of meat during storage. The statistical tests showed that the extract addition on active paper significantly affected the TVBN value ($P < 0.05$). Further test results showed that the treatment without the garlic extract (0%) was significantly different from the other treatments. However, the addition of 15% garlic extract was not significantly different from 20% garlic extract. The initial pH value of fresh meat before storage was 5.68 and classified as rotten after 9 days of storage for treatment without the extract. On the 15th day,

15% and 20% garlic extracts were put to active paper for beef. This is consistent with Ponnampalam et al. (2017) that the pH of normal meat ranges from 5.5 – 5.7, and meat with a pH of > 6 will be easily damaged by microorganisms experiencing some deterioration such as color and texture aroma.³⁵

An increase in the pH value of meat during storage indicates a decrease in quality, and anaerobic glycolysis lowers the pH to create lactic acid. The higher the lactic acid produced, the greater the decrease in the meat pH. This is consistent with Kiyimba et al. (2021), which stated that lactic acid production was the cause of the decrease in the pH of the glycolysis phase during postmortem.³⁶ The pH value of live meat ranges from 6.8-7.2, and after slaughter, it experienced a decrease in pH in the pre rigor condition of pH 7.2 to the ultimate pH between 5.4-5.8. The decrease stopped after the rigor mortise phase was reached in the meat. According to Leygonie et al. (2012), the decrease was also influenced by the ambient temperature (storage).³⁷ While storing meat at high temperatures accelerates the pace of pH decline, storing at low temperatures slows the rate of pH decrease.

3.4. Total Volatile Bases Nitrogen (TVBN)

Analysis of the TVBN value is one of the parameters used to determine meat freshness level. Figure 4 shows the changes in TVBN values in beef during cold storage.

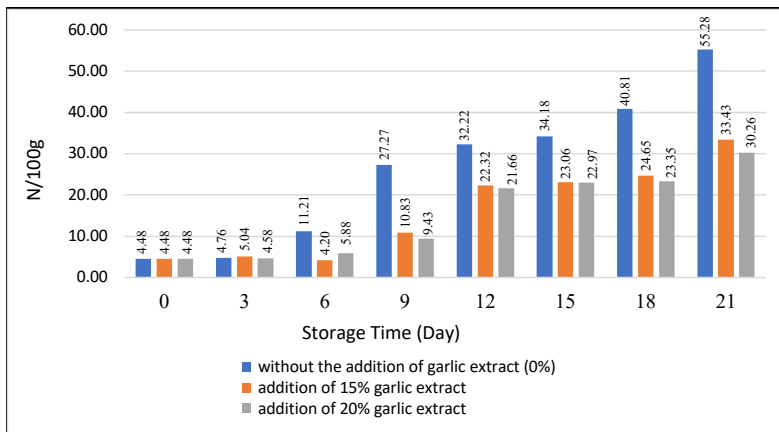


Figure 4. TVBN Value of Beef During Cold Storage

Based on statistical tests, the addition of garlic extract on active paper significantly affected the TVBN value ($P < 0.01$). Further test results showed that the treatment without the extract (0%) addition was significantly different from the other treatments. However, 15% garlic extract was not significantly different from the 20% extract. The TVBN value of meat on the first day of storage was 4.48 mg N/100, indicating a fresh category (Figure 4). On the 9th day of storage, meat treated without the extract had a TVBN value of 27.27 mg N/100g categorized as rotten. Meanwhile, the addition of 15% and 20% garlic extract with TVBN values of 22.32 mg N/100g and 21.66 mg N/100g had crossed the threshold on day 12 and was

categorized as rotten meat. TVB-N criteria for identifying rotten beef products are >20 mg/100 g according to the Ministry of Agriculture and Forestry of Korea, 2016³⁸. Based on the results obtained, the garlic extract affects the meat TVBN value during storage.

The increase in TVBN value is closely related to the decrease in beef quality. The treatment without onion extract reached the threshold value earlier due to the higher microbial activity in the sample. The high number of microbes in the sample breaks down more meat protein to produce volatile ammonia compounds. This is consistent with Holman et al. (2021), where increased microbial value degrades protein rapidly during storage. Furthermore, it increases the amount of ammonia and other volatile compounds, acting as indicators of decay and increasing the TVB value.³⁹ The increase in the TVBN value of beef with treatment without and with the garlic extract was also different due to the Allicin constituent and antimicrobial role. The inhibition of microbial growth minimizes the decomposition process of nutritional components in meat to reduce the TVBN value.

3.4. Total Bacteria of Total Plate Count (TPC) Method

The analysis determines the effect of using active paper on the total value of bacteria in meat during cold storage, as shown in Figure 5.

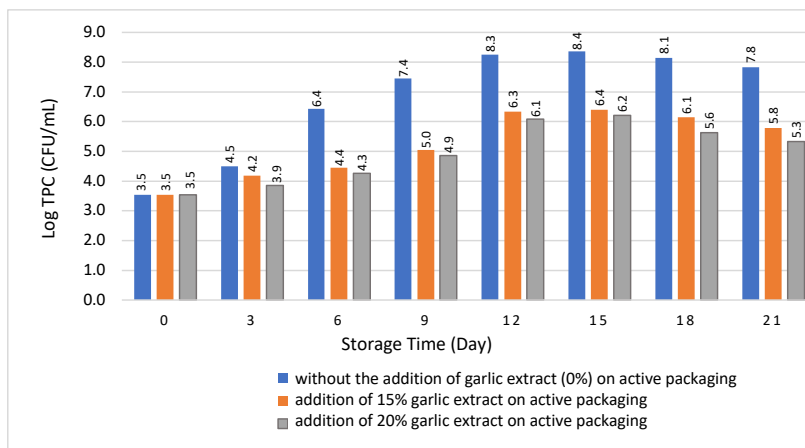


Figure 5. Graph of Beef TPC Value During Cold Storage

The garlic extract added to the active paper had a significant effect on the TPC value of meat ($P < 0.05$). Meanwhile, further test results showed that the treatment without garlic extract (0%) was significantly different from other treatments, but the addition of 15% was not significantly different from the 20% extract. The initial population of TPC meat was 3.5 log CFU/ml, categorized as fresh concerning microbiological quality (Figure 5). However, this value increased to 6.4 log CFU/ml on the 6th day of storage in the control sample, while the recommended maximum bacteriological limit was 6 log CFU/ml (SNI 3932:2008). The meat added with 15% and 20% garlic extract exceeded the maximum bacteriological limit on the 12th day of storage with TPC values of 6.3 log CFU/ml and 6.1 logs CFU/ml, respectively.

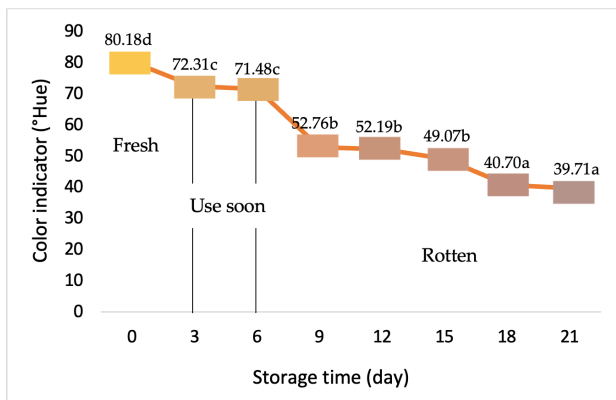
Therefore, the addition of 15% and 20% extract as active packaging inhibits microbial growth and extends the shelf life of meat up to 11 days longer than the samples without the garlic extract in the active packaging. This is because garlic contains an anti-microbial compound in the form of allicin.

Garlic extract applied to the activated paper diffuses to the entire surface of the beef to inhibit the growth of bacteria through total and partial inhibition of RNA, DNA, and protein synthesis. Furthermore, allicin blocks bacterial enzymes with thiol groups to ultimately inhibit bacterial growth. This is consistent with Deresse's (2010) statement that the mechanism of garlic inhibits bacterial growth by completely inhibiting RNA, DNA synthesis, and bacterial protein.⁴⁰ Mardiya (2018) also confirmed that the effectiveness of garlic extract with concentrations of 25%, 50%, and 100% was very effective in inhibiting *Staphylococcus aureus* bacteria and bactericidal. Meanwhile, the extract with a concentration of 12.5% was declared as an inhibitor of bacterial growth (bacteriostatic) but less effective at inhibiting bacteria (bacteriocidal).⁴¹

The storage temperature of beef can also affect the meat TPC value. Meat cells in the postmortem phase experience metabolic reactions, which are highly dependent on storage temperature. According to Comi (2017), the storage temperature and the metabolic reactions in meat are inversely proportional.⁴² This is supported by Kuswandi and Nurfawaidi (2017), where beef stored at room temperature is categorized as unfit for consumption at 10 hours with a TPC value of 6,711 CFU/mL. Meanwhile, at cold temperature, it is categorized as unfit for consumption on day 7 with a TPC value of 6961 CFU/mL.²⁹

3.5. Color Change Response on Smart Indicator Labels for Beef Packaged in Active Packaging at Cold Temperature Storage

The color change analysis of the smart indicator determines the effect of adding garlic extract to active packaging with different concentrations on beef during storage at cold temperatures (chiller). The color change of the resulting indicator label is information on the freshness condition of the packaged beef. Significant color changes on indicators ease consumers to measure the freshness level based on visual appearance, as seen in Figure 6.



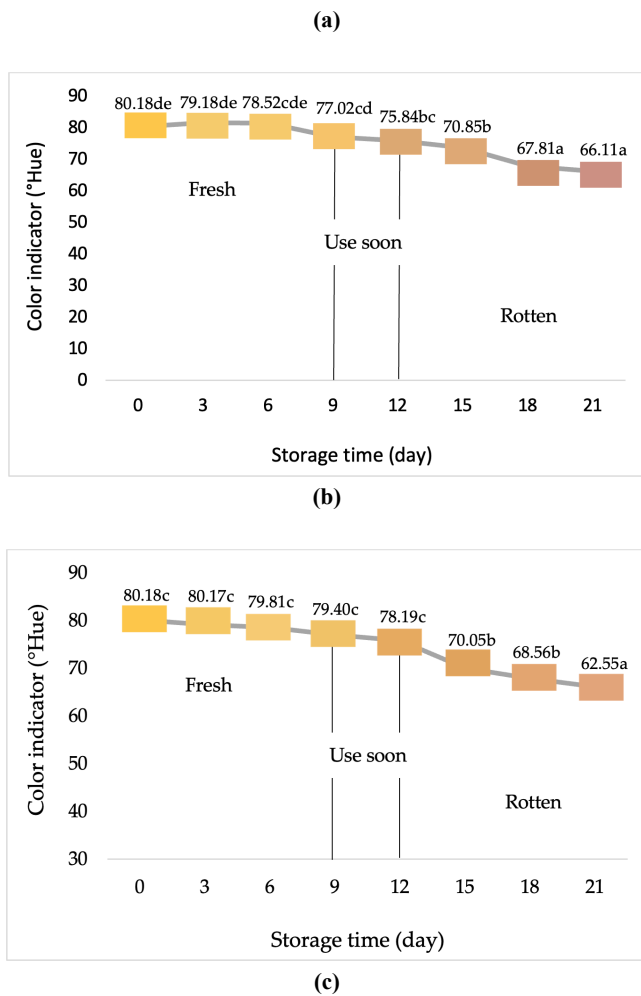


Figure 6. Color Change Profile of Smart Indicator Labels with Garlic Extract Addition on the Concentrations at 0% (a), 15% (b), and 20% (c)

Figure 6 showed that the °Hue value on the smart indicator label decreased during storage, from dark yellow to faded red. The color change is a response to the quality condition. There are three phases of change on smart packaging labels in the description. These include phase I with dark yellow indicating fresh, phase II with yellow and red gradations indicating immediate consumption, and phase III with faded red indicating damaged meat unfit for consumption.

The color change on the smart indicator label is closely related to the increased number of microbes and enzymes in beef during storage. These changes are caused by the

decomposition of nutritional components of meat by enzymes and microbes, producing volatile base compounds used as an early sign of damage in packaged beef. This is consistent with Sani et al. (2021) that volatile ammonia compounds in meat can increase the pH.⁴³ The increase in the pH of meat is caused by ammonia reacting with H⁺ ions to produce H ions. The OH⁻ ions in the packaging are directly proportional to the pH value, affecting the change in smart indicator labels.

3.6. Correlation of Color Value Changes in Smart Packaging Indicators BTB: PR (1:1) with the Effect of Active Packaging on All Meat Spoilage Parameters

The relationship between the color change response of smart indicator labels with all test parameters on beef, such as TVBN, TPC, and pH, determined the sensitivity of smart indicator labels in detecting quality deterioration. This obtains synchronization between changes in the color of the smart indicator label to the deterioration of beef quality.

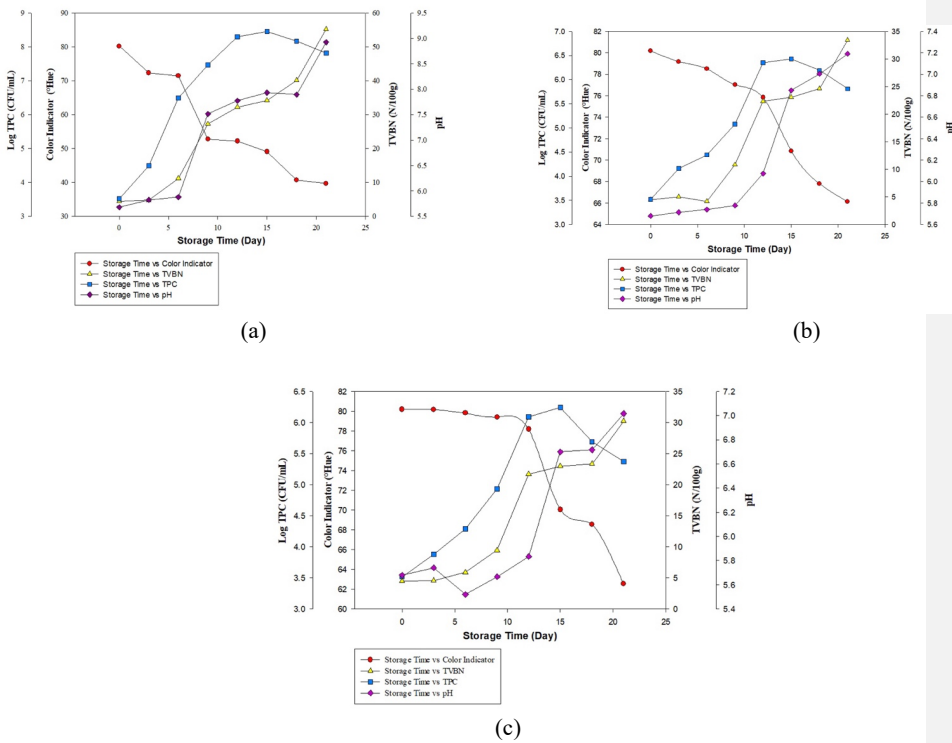


Figure 7. Correlation of Color Change on Smart Indicator Label with All Beef Spoilage Parameters with the Garlic Extract Addition (a) Control, (b) 15%, (c) 20%

Based on Figure 7, beef stored without the garlic extract (0%) showed that the increase in the TVBN value and the pH of the meat was in line with the decrease in the color value of

the smart packaging indicator. However, it is not consistent with the results of the TPC value since the beef TPC value has crossed the threshold on the 6th day of storage, which is 6.4 CFU/ml. Meanwhile, the TVBN value is suitable for consumption on the 6th day. Mansur et al. (2016) also reported that the TVB-N values were inconsistent with the viable microbial count (TVC). Based on TVB-N, the samples were classified as damaged after seven and eleven days of storage.⁴⁴ This is because the microbial calculation method is not selective against spoilage bacteria in meat but counts all the pathogenic microbes. Holman et al. (2021) also stated that the TVB-N value was more precise and accurate in classifying beef in fresh or rotten conditions than the total microbial value (TVC).³⁹

Based on Figures 7(b) and (c), beef stored with 15 % and 20% garlic extract treatment showed that the increase in the parameters of the meat quality deterioration, including TPC, TVBN, and pH, was consistent with the decrease in the color of the smart indicator label. The smart indicator changes color from yellow (fresh) to faint red (rotten) during storage. The increase in TVBN, TPC, and pH values is influenced by the high nutritional content, specifically protein broken down into polypeptides and other amino acids through the deamination process to form ammonia. With the increase in microbes, the nutritional components decomposed by microbes are also greater and produce basic volatile compounds increasing the pH value of meat during storage. The resulting volatile compounds accumulate in the packaging and are directly detected by the smart indicator label. The reduction in the color value of the smart indicator label shows the presence of many alkaline volatile chemicals in the package and the rottenness and unfitness for ingestion of the meat. Furthermore, when viewed from the visual appearance on the 9th and 12th day of storage without (0%) and with (15% and 20%) garlic extract, the beef was damaged and unfit for consumption characterized by the formation of mucus, emits a sour/rotten smell, and the texture begins to be sticky to the touch.

The increase in beef quality deterioration parameter value for each treatment was different due to the addition of garlic extract as active packaging. The extract has a bioactive compound in allicin to inhibit microbial growth. This compound in the package diffuses in the meat surface to suppress the microbe growth. According to Deresse (2010), it works by completely inhibiting bacterial RNA synthesis and inhibiting DNA and protein synthesis.⁴⁰ The pattern of relationship between the parameters of the decline in beef quality and the color analysis value of indicator labels is also consistent with Kuswandi and Nurfawaidi (2017), where a linear relationship pattern to the parameters of the beef freshness level with changes in the color intensity of the bromocresol purple and methyl red smart labels were observed.²⁹ In this study, beef treated without the garlic extract experienced a decrease in quality and was not suitable for consumption on the 9th day. Meanwhile, with 15% and 20% garlic extract, the meat was damaged and unfit for consumption on the 12th day of storage. This indicates that garlic extract storing at cold temperatures can extend the shelf life because of the Allicin properties. Visually, the smart indicator label using Whatman paper no. 1 as the base material and the BTB + PR (pH 5.00) used in smart packaging is used for beef freshness with a color change from yellow (beginning) to faint red (late). Therefore, it assists customers in determining freshness without touching the texture or opening the packaging.

4. Conclusion

The garlic extract in active packaging, supported by the microbiological evaluation results of TVBN and pH, extended shelf life during storage. In addition, the results showed that the active paper with 15% and 20% garlic extract extended the shelf life for 12 days of cold storage. The color change profile of the smart indicator label combination of BTB + PR pH 5.00 was easily observed from dark (fresh) to reddish yellow (to be consumed immediately). The red color fades when the beef is rotten and unfit for consumption. Therefore, the combination of active and smart packaging is suggested as a potential packaging application in fresh meat packaging.

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Author(s)

Andi Dirpan

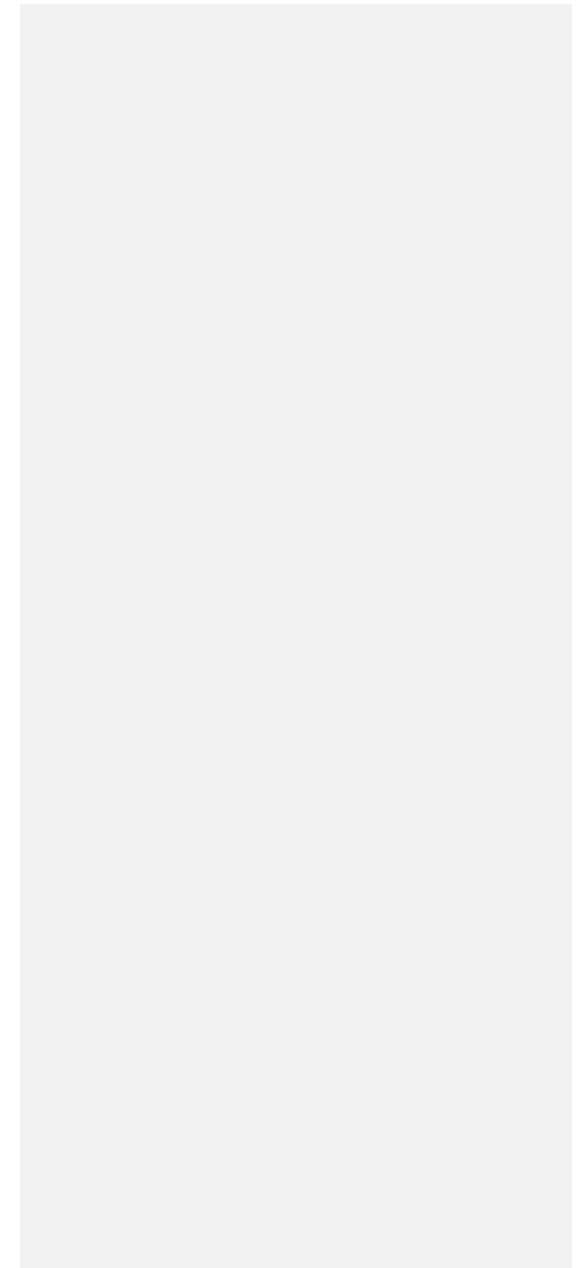
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Authors: Andi Dirpan *, Muspirah Djalal, Andi Fadiah Ainani
Received: 30 April 2022
E-mails: dirpan@unhas.ac.id, muspirah_djalal@agri.unhas.ac.id, andifadiah97@gmail.com

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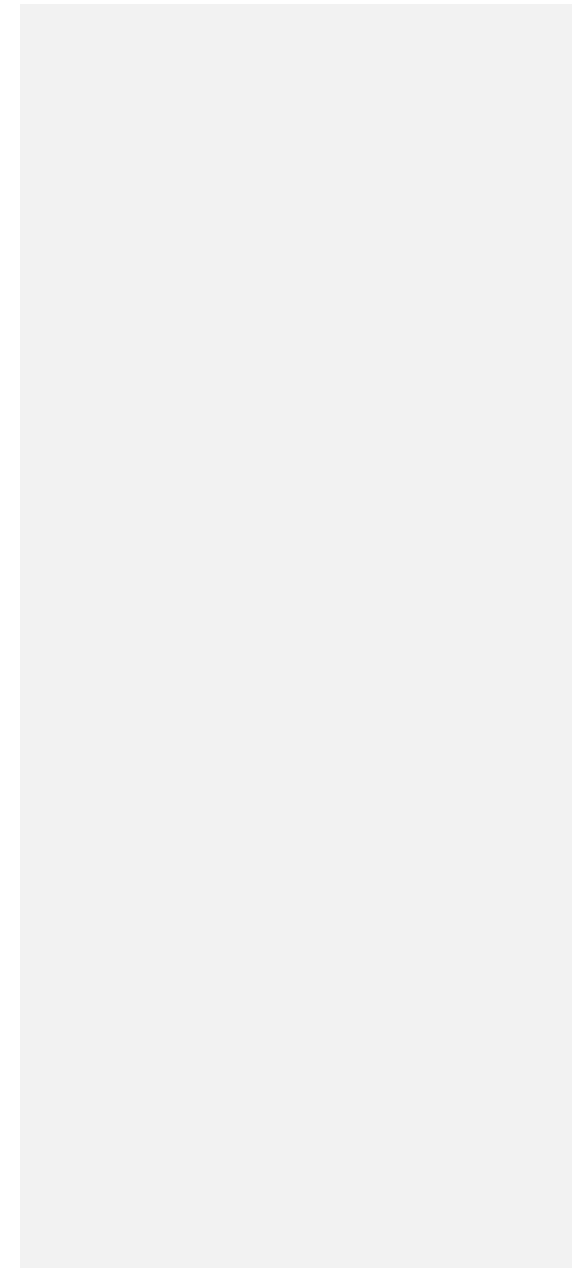
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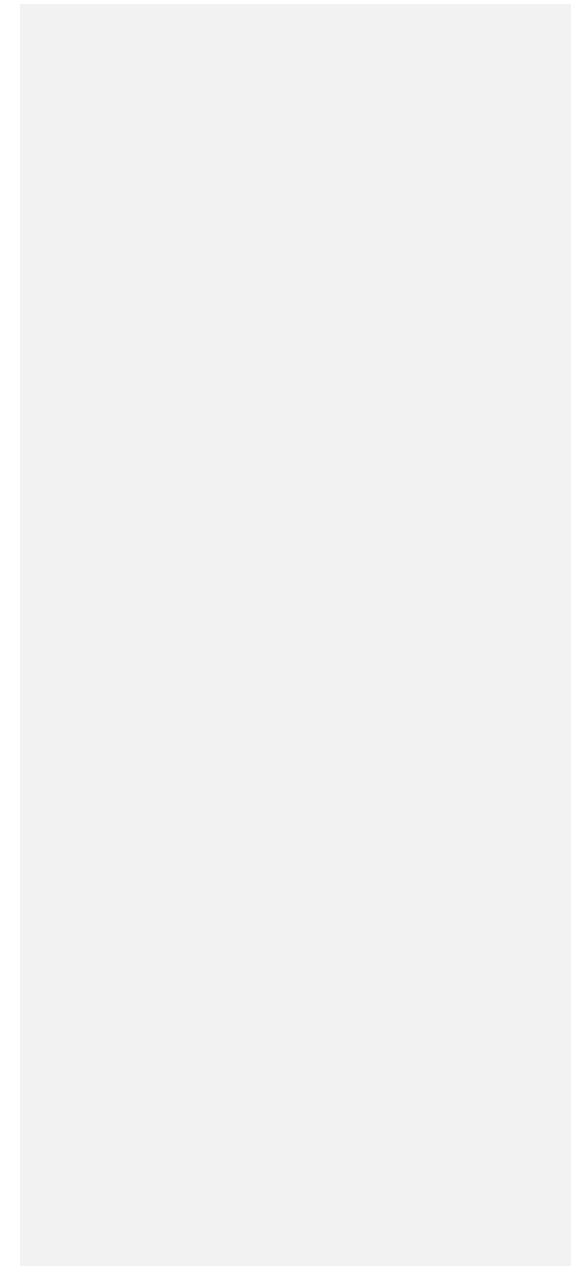
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-Revisions and Amends

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- Manuscript revisions





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Authors: Andi Dirpan *, Muspirah Djalal, Andi Fadiah Ainani

Received: 30 April 2022

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Authors: Andi Dirpan *, Muspirah Djalal, Andi Fadiyah Ainani

Received: 30 April 2022

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Abstract

Safety and quality, as the major concerns of meat, are highly dependent on the ingredients and packaging techniques used. A basic combination of active and intelligent packaging is believed to be capable of preserving product quality, extending shelf life, and monitoring product deterioration. Therefore, this study aimed to extend and monitor the beef quality at cold temperatures (4 ± 1 °C). The active packaging applied garlic extract (0%, 15%, and 20% (w/w)) to release anti-microbial agents. Meanwhile, the intelligent paper applied a combination of bromothymol blue (BTB) and phenol red (PR) solutions at pH 5.00. The results showed that beef packed without the addition of garlic extract had already deteriorated on the 6th day of storage while, with the addition of garlic extract (15% and 20%) rotted on the 12th day. The intelligent indication label's color profile changed from dark yellow (fresh), to reddish-yellow (to be consumed immediately), to faded red (rotten). The color change of the intelligent indicator label in response to all meat deterioration criteria demonstrates a linear correlation for determining the extent of rottenness during storage. Therefore, this simple combination of active paper and intelligent indicator can be used to extend the shelf life and monitor meat quality.

Keywords

smart packaging; meat; garlic oil; shelf life

Review Report Form

English language and style

- Extensive editing of English language and style required
- Moderate English changes required
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	Yes	Can be improved	Must be improved	Not applicable
Does the introduction provide sufficient background and include all relevant references?	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
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- Regarding the packaging structure, shown in Figure 1, Styrofoam is a polystyrene foam whereas the authors indicate that a polypropylene tray is used. The procedure was probably not expressed correctly. Moreover, all the packaging films or products should be included in the Materials section, indicating the commercial reference and main characteristics. Furthermore, the use of PS, LDPE or PP is not sustainable and it is suggested to, at least, indicate that those should be replaced in future developments.
- The organization of the Results section can be improved. For instance, the article can first present results in relation with the quality parameters of meat (pH, color, microbial counts, etc.) and then effect of the additives on the antimicrobial properties and color (active and intelligent properties)
- Results should be discussed and compared with other studies reporting the shelf-life stability of meat packaged. Please see for instance 10.3390/foods11030426
- The potential effect of the garlic extract in the organoleptic properties of meat should be discussed in the Conclusions

Submission Date: 30 April 2022

Date of this review: 12 May 2022 13:51:15

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MATRICES OF AMENDMENTS FOR REVIEWER 1

Comments and Suggestions for Authors	Author's responds
Regarding the packaging structure, shown in Figure 1, Styrofoam is a polystyrene foam whereas the authors indicate that a polypropylene tray is used. The procedure was probably not expressed correctly. Moreover, all the packaging films or products should be included in the Materials section, indicating the commercial reference and main characteristics. Furthermore, the use of PS, LDPE or PP is not sustainable and it is suggested to, at least, indicate that those should be replaced in future developments.	Thank you for your comments, For the material we used to transfer the meat from the slaughterhouse to the lab we used polypropylene and for the tray in the packaging system we used Styrofoam (polystyrene), so it is actually for a different purpose. As suggested, all the materials have been included in the materials section and the main characteristics also have been added in line 103-108. For the use of PS and LDPE yes, it is certainly not sustainable and as suggested we already put a recommendation to replace it in future in line 208, thank you.
The organization of the Results section can be improved. For instance, the article can first present results in relation with the quality parameters of meat (pH, color, microbial counts, etc.) and then effect of the additives on the antimicrobial properties and color (active and intelligent properties)	Thank you for your input. We have rearranged the results
Results should be discussed and compared with other studies reporting the shelf-life stability of meat packaged. Please see for instance 10.3390/foods11030426	Thank you for your comment. The result has been compared with other studies and the literature suggested also added in the method.

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Topic : [Smart Technologies in Food Packaging and Sensors](#)

Abstract

Safety and quality, as the major concerns of meat, are highly dependent on the ingredients and packaging techniques used. A basic combination of active and intelligent packaging is believed to be capable of preserving product quality, extending shelf life, and monitoring product deterioration. Therefore, this study aimed to extend and monitor the beef quality at cold temperatures (4 ± 1 °C). The active packaging applied garlic extract (0%, 15%, and 20% (w/w)) to release anti-microbial agents. Meanwhile, the intelligent paper applied a combination of bromothymol blue (BTB) and phenol red (PR) solutions at pH 5.00. The results showed that beef packed without the addition of garlic extract had already deteriorated on the 6th day of storage while, with the addition of garlic extract (15% and 20%) rotted on the 12th day. The intelligent indication label's color profile changed from dark yellow (fresh), to reddish-yellow (to be consumed immediately), to faded red (rotten). The color change of the intelligent indicator label in response to all meat deterioration criteria demonstrates a linear correlation for determining the extent of rottenness during storage. Therefore, this simple combination of active paper and intelligent indicator can be used to extend the shelf life and monitor meat quality.

Keywords

smart packaging; meat; garlic oil; shelf life

Review Report Form

English language and style

- Extensive editing of English language and style required
 Moderate English changes required
 English language and style are fine/minor spell check required
 I don't feel qualified to judge about the English language and style

	Yes	Can be improved	Must be improved	Not applicable
Does the introduction provide sufficient background and include all relevant references?	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Are all the cited references relevant to the research?	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Is the research design appropriate?	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Are the methods adequately described?	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Are the results clearly presented?	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Are the conclusions supported by the results?	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Comments and Suggestions for Authors

The work concerns an important aspect of food storage and ensuring food safety. In general, the work is written correctly and, after introducing minor corrections.

Line 62 there is "... its volatile Allicin constituent"... should be "...its volatile allicin constituent"...

Line 107-112 This description is not clear. How can blended garlic be immersed in alcohol for several times. The mixing process is not imprecisely described. What was the speed of mixing, was it a mechanical stirrer?

Line 118 Are you sure that tapioca starch was dissolved? Was it soluble starch and real solution or a suspension of starch in water.

Line 122 there is "*tween 80*" should be "*Tween® 80*"

Line 123 "The paper dough is poured" should be "The paper dough was poured"

Line 126 "surface is pressed" should be "surface was pressed"

Line 235 "quantification data will be compared" should be "quantification data was compared"

Please check the description of the methodology in terms of grammar.

Table 1 In my opinion, it does not make sense to give the average pH value, especially in relation to the average pH on the given day (last column). These are different samples, so what would such an average be evidence of? The average in the last line makes mathematical sense, but it might be better to present the difference from the initial pH.

I have the same comments about table 2 and 3.

I have some concerns about the color indicator. Is it certain that the consumer is able to distinguish borderline indicator colors? The colors presented in the charts are quite similar in the borderline ranges. Perhaps the total color difference (delta E) should be determined. For this parameter, the limit values are known which make it possible to conclude whether the consumer will see a color difference or not. Given the results from the CieLab system, it is easy to calculate. Alternatively, a consumer test can be carried out.

Submission Date: 30 April 2022

Date of this review: 11 May 2022 11:46:50

MATRICES OF AMENDMENTS FOR REVIEWER 2

Comments and Suggestions for Authors	Author's responds
Line 62 there is "... its volatile Allicin constituent"... should be "...its volatile allicin constituent"...	Thank you for your response and we greatly appreciate it. We have changed all capital into lowercase except for the one at the beginning of the sentence.
Line 107-112 This description is not clear. How can blended garlic be immersed in alcohol for several times. The mixing process is not imprecisely described. What was the speed of mixing, was it a mechanical stirrer?	Thank you for your detail comment. We are sorry that we create confuse with the way we write the sentence. The 3x24 h actually means 72 hours and not a multiple times immersion. It is already written 72 hours in the paper in line 114. While for the stirring process, we understand that the sentence creates confusion. It is actually a manual gentle stirring process in which during the 72 hours of immersion process the solution was stirred every 5 hours. The changed was made in line 114-115
Line 118 Are you sure that tapioca starch was dissolved? Was it soluble starch and real solution or a suspension of starch in water.	Thank you for the correction. You are right it is suspension so we have changed it into suspended in line 123 instead of dissolved.
Line 122 there is "tween 80" should be "Tween® 80"	Thank you for the correction. The change has been made in line 128 according to your suggestion.
Line 123 "The paper dough is poured" should be "The paper dough was poured"	Thank you for the correction. The change has been made in line 130 according to your suggestion
Line 126 "surface is pressed" should be "surface was pressed"	Thank you for the correction. The change has been made in line 132 according to your suggestion
Line 235 "quantification data will be compared" should be "quantification data was compared"	Thank you for the correction. The change has been made in line 308 according to your suggestion
Please check the description of the methodology in terms of grammar.	Thank you for your comments. We rechecked the methodology and edit several parts to make it more readable. We hope this meets your expectation.
Table 1 In my opinion, it does not make sense to give the average pH value, especially in relation to the average pH on	Thank you for your suggestion. We appreciate your input and we have deleted the last column.

<p>the given day (last column). These are different samples, so what would such an average be evidence of? The average in the last line makes mathematical sense, but it might be better to present the difference from the initial pH.</p>	
<p>I have the same comments about table 2 and 3.</p>	<p>Thank you for your suggestion. We appreciate your input and we have deleted the last column.</p>
<p>I have some concerns about the color indicator. Is it certain that the consumer is able to distinguish borderline indicator colors? The colors presented in the charts are quite similar in the borderline ranges. Perhaps the total color difference (delta E) should be determined. For this parameter, the limit values are known which make it possible to conclude whether the consumer will see a color difference or not. Given the results from the Cielab system, it is easy to calculate. Alternatively, a consumer test can be carried out.</p>	<p>Thank you for your comments, it very much enlightened us. We added the Delta E value and also an explanation of this value is the result of our study.</p>

Article

A Simple Combination of Active and Intelligent Packaging Based on Garlic Extract and Indicator Solution in Extending and Monitoring the Meat Quality Stored at Cold Temperature

Andi Dirpan *, Muspirah Djalal and Andi Fadiyah Ainani

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* Correspondence: dirpan@unhas.ac.id

Abstract: Safety and quality, as the major concerns of meat, are highly dependent on the ingredients and packaging techniques used. A basic combination of active and intelligent packaging is believed to be capable of preserving product quality, extending shelf life, and monitoring product deterioration. Therefore, this study aimed to extend and monitor the beef quality at cold temperatures (4 ± 1 °C). The active packaging applied garlic extract (0%, 15%, and 20% (*w/w*)) to release antimicrobial agents. Meanwhile, the intelligent paper applied a combination of bromothymol blue (BTB) and phenol red (PR) solutions at pH 5.00. The results showed that beef packed without the addition of garlic extract had already deteriorated on the 6th day of storage while, and with the addition of garlic extract (15% and 20%) rotted on the 12th day. The intelligent indication label's color profile changed from dark yellow (fresh), to reddish-yellow (to be consumed immediately); to faded red (rotten). The color change of the intelligent indicator label in response to all meat deterioration criteria demonstrates a linear correlation for determining the extent of rottenness during storage. Therefore, this simple combination of active paper and intelligent indicator can be used to extend the shelf life and monitor meat quality.

Keywords: smart packaging; meat; garlic oil; shelf life

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1. Introduction

Meat is one of the most essential nutrients, containing high levels of easily digestible protein, calorie-dense fat, vitamins, and other micronutrients. Its high nutritional value corresponds to its rising demand as the world's most significant food product during the past several years [1]. The high demand should be fulfilled with the availability and an increase in the fresh meat grade, including the quality and safety, since beef is a perishable commodity [2]. The decrease in quality is due to the high fat and water content, making it susceptible to microbial contamination and lipid oxidation.

Proper packaging plays a critical role in preserving the quality and safety of meat. It is critical to select materials that will preserve the packaged product's qualities and assure its safety. The use of active packaging in conjunction with cold storage is one of the efforts that can be made to prolong the shelf life and enhance the quality.

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Active packaging applications can be used to inhibit microbial growth in meat [3–8]. These applications incorporate specific active substances into packaging methods to preserve food products by inhibiting bacteria contaminations that contribute to spoilage during storage [9]. The spoilage inhibition mechanism can be conducted in two ways, namely direct and indirect contact. In the direct contact, the packaging materials will preserve food through direct contact, while in the indirect contact, the active packaging releases volatile anti-microbial agents to the packaging system where the food product is placed [10,11]. Camo et al. and Nerin et al. employed an active packaging technique incorporating rosemary and oregano directly touching the meat [12,13], and Campos-Requena et al. placed active packaging containing essential oil to strawberries that was not in direct contact with them [14], and both resulted in a synergistic antibacterial activity.

Volatile anti-microbial compounds are essential oils derived from herbs and spices, such as oregano, thymol, carvacrol, cinnamon, rosemary, ginger, lemongrass, and garlic [15]. Garlic (*Allium sativum*) contains the antibacterial component allicin, which is active against both Gram-positive and Gram-negative bacteria [16,17]. The anti-microbial activities of different species of *Allium* have been investigated [18–21]. Several studies have been conducted on the administration of garlic in active packaging, including the following: Zheng Dong et al. developed an active packaging material based on a PP/LDPE composite film containing *Allium sativum* essential oil. The addition of essential oils into the packaging extends the shelf life of large yellow croaker (*Pseudosciaena crocea*) fillets for 5 days stored at 4 ± 1 °C [22]. Seydim and Sarikus also reported that the anti-microbial properties of whey protein isolate containing essential garlic oil with a ratio of 3.0–4.0% (wt/vol) effectively inhibited the growth of *Salmonella enteritidis*, *S. aureus*, *E. coli* O157:H7, *L. monocytogenes*, and *Lactobacillus plantarum* [23]. The use of garlic extract in active packaging can inhibit microbial growth due to its volatile allicin constituent [24]. Allicin chemicals suppress microbial development by increasing their permeability in penetrating bacterial cell walls to decrease protease enzyme production and disrupt protein and nucleic acid metabolism [19].

The application of active packaging on fresh meat can be accompanied by the addition of intelligent packaging in the form of indicators that monitor changes in quality. This is achieved through chemical reactions between indicators and the results of microbial metabolism or changes in the meat's chemical composition. Intelligent packaging incorporates indicators that convey information about the product's quality without causing damage to the packaging and not also in food products. This reduces the risk of loss due to product damage and provides a more accurate condition estimate than expired labels [25,26]. Furthermore, intelligent packaging also assists distributors in adjusting the prices of the products sold [27]. For example, lowering prices when quality has decreased to avoid losing food products. Previous studies applied this method to avocados [28], tuna fillets [29], shrimp [30], cherry tomatoes [31], and *arummanis* mango [32]. Monitoring of color changes can be done due to differences in pH of fresh product and damaged product. According to Shukla et al., color-based pH indicators can be used as intelligent packaging to monitor the presence of volatile amines from microbial metabolites in meat spoilage [33]. The increase in pH during decay causes a significant change in the indicator's color to be

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visually observed. Meanwhile, Romero et al. evaluated the quality of packaged cow's milk using two intelligent packaging prototypes based on the bromothymol blue pH indicator [34]. Julyaningsih et al. and Yolanda et al. combined active and intelligent packaging based on pH indicators methyl red and bromothymol blue to monitor the tuna fillet's quality [35,36]. Meanwhile, Yong and Liu combined anthocyanin-rich extracts into biopolymer-based films to extend shelf life and monitor the food product's quality [37].

The spoilage and contamination of food supplies jeopardize global food security. Therefore, the innovative application of active and intelligent packaging needs to be developed. As a result of these considerations, the purpose of this research is to evaluate the efficacy of active paper with garlic extract addition when applied to fresh beef during storage at low temperatures (4 ± 1 °C). Furthermore, it investigates the relationship (correlation) between color analysis of intelligent packaging indicators on various parameters of the beef spoilage test.

2. Materials and Methods

2.1. Materials

The beef tenderloin was purchased fresh at the slaughterhouse, Tamangapa Raya Antang. Meanwhile, garlic, as an anti-microbial compound in the activated paper, was purchased from supermarkets. Bromothymol blue (Merck, Darmstadt, Germany, CAS No: 76-59-5) and phenol red (Merck, Darmstadt, Germany, CAS No. 34487-61-1) color indicators in intelligent packaging were purchased from Sigma-Aldrich. Mueller Hinton Agar (Oxoid, CM0405) and Nutrient Agar (Merck, Darmstadt, Germany). This research also uses *Escherichia coli* (ATCC 25922), *Staphylococcus aureus* (ATCC 29213), *Pseudomonas aeruginosa* (ATCC 27853), chitosan food grade (YXchuang, Cina), Tween® 80 (Merck, Darmstadt, Germany), corn starch, 96% glacial acetic acid (Brenntag Inc., Germany), potassium carbonate (K_2CO_3) (Merck, Darmstadt, Germany), 7% trichloroacetic acid (TCA) (Merck, Darmstadt, Germany), hydrochloric acid (HCl) (Merck, Darmstadt, Germany), and 0.85% sodium chloride (NaCl) (Merck KGaA, Germany).

2.2. Garlic Extract Preparation

Fifty grams of garlic was blended until smooth and put into a container containing 250 mL of 96% alcohol for a 72 h immersion. During the immersion time, the solution was gently stirred 14 times or every 5 h. Subsequently, the garlic dregs and the filtrate were separated by filtering. At 45 °C, the filtrate was evaporated with an industrial rotary evaporator (TMAX-2L-1) until it became thick reddish-yellow garlic [38].

2.3. Active Paper Preparation

Active paper preparation was carried out by cutting Whatman paper no. 1 into small pieces, then 15 g of the small pieces paper was soaked in 250 mL of distilled water for 24 h. The soaked paper was added with 250 mL of distilled water and crushed using a blender for 20 min until a pulp was formed. The paper pulp obtained was squeezed to remove the water, meanwhile 30% (*w/w*) tapioca starch was suspended in 50 mL of distilled water, then the pulp and tapioca suspension were homogenized. Furthermore, 100 mL of acetic acid (1%) containing 0.45% chitosan was

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added to the tapioca and pulp mixture and homogenized with a blender for 5 min. Garlic extract with a concentration according to the treatment (0%, 15% and 20%) (*w/w*) was added to the mixture; subsequently, 50 mL of distilled water was also added to the mixture. Furthermore, 0.205 g of Tween® 80 was added and mixed at room temperature using a hotplate magnetic stirrer to generate an emulsion. The paper dough was poured over the surface of the printer container following the styrofoam size used as a beef storage container and flattened to make a wet paper sheet. ~~After that~~, the wet paper sheet was pressed between the glass surfaces with a load of 2 kg. It was then dried at 40 °C for 48 h, and during the drying process the paper was turned over after 24 h of drying (Modification of Wiastuti et al. [39]).

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2.4. Indicator Solution Preparation

BTB and PR (1:1) indicator solutions were made by dissolving 1% (*w/v*) of the solution in 95% ethanol. Furthermore, the indicator was adjusted to pH 5 using a solution of glacial acetic acid [40].

2.5. Intelligent Indicator Label Preparation

Whatman paper no. 1 was cut with a size of 2 × 4 cm and immersed in 20 mL of indicator solution for 12 h at room temperature (28 ± 2 °C). The indicator labels were ~~then~~ dried using an electric dryer (Philips PH 8102) and ~~in~~ a closed container with a drying distance of 30 cm.

2.6. Application of Active and Intelligent Packaging on Fresh Beef

Fresh beef (tenderloin) with normal pH (5.6–5.7) was obtained from the Tamangapa Antang Makassar abattoir, which was taken at a relative postmortem time of about 3 h. The meat was packaged plastic (PP) containers and put in a cooler box, then immediately transported to the laboratory and prepared sterilely into 180-g pieces/packages. Activated paper with different concentrations was inserted into the styrofoam base. However, since the application of plastic is not sustainable, it is suggested to use more sustainable materials in future. The pieces of meat were packed in styrofoam (1.05 g/cm³) and covered with active paper ~~according to the treatment (0%, 15% and 20% of garlic extract)~~ that filled the entire base. The indicator label was placed inside the package by sticking to the LDPE film surface (0.9 g/cm³) used as a cover. Observation of beef (tenderloin) packaged with active and intelligent packaging was conducted at a cold temperature (chiller) of 4 ± 1 °C every 3 × 24 h for 21 days of storage. The design for implementing active and intelligent packaging is presented in Figure 1.

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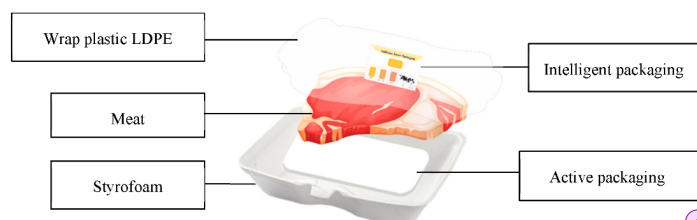


Figure 1. Design of the active packaging and intelligent system.

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2.7. Antibacterial Activity Testing of Agar Diffusion Method on Activated Paper

The antibacterial activity of active paper was tested to determine the inhibition toward the growth of Gram-positive bacteria *Staphylococcus aureus*, and Gram-negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa*. Sheets of active paper with a diameter of 5.5 mm were placed on Mueller Hinton Agar (MHA) media, which spread 0.1 mL of the microorganism culture on its surface. Furthermore, the Petri dishes were incubated at 37 °C for 24 h, and after incubation, a clear zone (inhibition zone) was formed around the active paper. The resistance diameter was measured in millimeters (mm) using a caliper [41], and each test was conducted in 3 replications to obtain the average result.

2.8. Parameters of Beef Observation Packaged with a Combination of Active and Intelligent Packaging

2.8.1. pH Measurement

The initial and final pH of fresh beef during storage were measured using Horiba Laquatwin Compact pH Meter P-33 with an accuracy of 0.01%. A total of 1 g of the sample was mashed and dissolved in 10 mL of demineralized water. Furthermore, it was inserted on the surface of the pH meter sensor until the value was ~~displayed~~ on the pH meter screen.

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2.8.2. TVBN Value Analysis

The meat sample was weighed at 1 g ± 0.1 g, pulverized in a mortar with 3 mL of 7% TCA solution, and filtered to obtain the filtrate. The boric acid solution of 1 mL was put into the “inner chamber” of the conway cup, and the lid was placed in a position to cover the cup. Furthermore, the filtrate was fed into the outer chamber on the left. Then 1 mL of saturated K₂CO₃ solution was added to the outer chamber on the right, ensuring that the filtrate and K₂CO₃ do not mix. The cup was instantly closed and rotated to mix the two liquids in the outer chamber. The blank solution was prepared the same way as the standard solution, except that the filtrate was replaced with a 7% TCA solution. Subsequently, it was stored at 28 ± 2 °C for 24 h, and the boric acid solution in the inner chamber containing the blank was added with two drops of color indicator solution, then titrated with 0.02 N HCl until it turned pink. The conway cup containing the filtrate was added with two drops of color indicator solution and titrated with the same solution until it turned pink (SNI-2354.8:2009; AOAC, 1995). Equation (1) for determining TVBN:

$$\text{TVBN level (mg/100 g)} = \frac{(V_c - V_b) \times N \times 14.007 \times 100}{w}$$

Description:

V_c = volume of HCl solution in sample titration

V_b = volume of HCl solution in blank titration

N = normality of HCl solution

W = sample weight (g)

14.007 = atomic weight of nitrogen

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2.8.3. Calculation of Total Bacterial Count

Total bacteria were calculated based on total plate count method (SNI 2897:2008). The sample solution was made from 1 g of meat homogenized with 9 mL of sterile physiological solution (0.85% NaCl), referred to as the 10^{-1} dilution. Then, the serial dilution was made up to a concentration of 10^{-6} . Microbial cultivation using the pour technique in which 1 mL dilutions of 10^{-4} , 10^{-5} , and 10^{-6} were put into separate sterile petri dishes in duplicate. Subsequently, ± 15 mL of sterile NA media was added into the petri dishes, then it was homogenized and incubated at 30°C for 48 h. Equation (2) for calculation of the total plate count:

$$N = \frac{\Sigma C}{[(1 \times n_1) + (0.1 \times n_2) + \dots] \times (D)} \quad (2)$$

Description:

N = number of colonies per mL/per gram of product

ΣC = total number of colonies counted

n_1 = number of cups in the first dilution

n_2 = number of cups in the second dilution

D = first dilution calculated

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2.9. Intelligent Indicator Color Measurement Quantification

The color measurement of the intelligent indicator label was analyzed using a digital color meter (T-135). This quantification is conducted by attaching the colorimeter sensor to the intelligent indicator label. The tool will then show the values of L (lightness), a (redness), and b (yellowness) on display. These three values are international standards of color measurement published by the Hunterlab Association Laboratory (2008). Furthermore, the intelligent packaging indicator color is determined by calculating the $^{\circ}\text{Hue}$ value using the Formula (3), and the difference of total color (ΔE) was measured with this Formula (4) [42,43]:

$$^{\circ}\text{Hue} = \tan^{-1} \frac{b}{a} \quad (3)$$

Description:

$^{\circ}\text{Hue}$ = parameters for color range

a = is a red-green mixed color

b = is a red-green mixed color yellow-blue

$$\Delta E^* = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2} \quad (4)$$

2.10. Level of Relationship (Correlation) between Test Parameters

The intelligent packaging indicator quantification data was compared with the data for each beef spoilage parameter presented in a graph using the Sigma Plot software version 14.0.

2.11. Statistical Analysis

All data from the test parameters were analyzed using analysis of variance (ANOVA) with Duncan's multiple range test using SPSS software version 22.0 (IBM Corp., United States), and the calculated values differed significantly when $p < 0.05$.

3. Results and Discussion

3.1. Meat pH

The degree of acidity (pH) in meat is one measure of its freshness, therefore, the analysis of the meat pH value was performed to determine the effect of adding various concentrations of garlic extract during cold storage. Table 1 shows the pH changes that occur in beef during storage.

Table 1. The pH value of packaged meat samples stored in cold storage for 21 days.

Storage Time (Day)	Addition of Garlic Extract on Active Paper		
	0%	15%	20%
0	5.68 ± 0.06 g	5.68 ± 0.06 g	5.68 ± 0.06 g
3	5.82 ± 0.16 g	5.71 ± 0.01 g	5.74 ± 0.16 g
6	5.88 ± 0.0 g	5.74 ± 0.01 g	5.52 ± 0.16 g
9	7.52 ± 0.72 ^{b,c,d}	5.78 ± 0.07 g	5.67 ± 0.16 g
12	7.77 ± 0.52 ^{b,c}	6.08 ± 0.01 ^{f,g}	5.83 ± 0.13 g
15	7.93 ± 0.20 ^b	6.85 ± 1.27 ^{d,e,f}	6.70 ± 0.62 ^{e,f}
18	7.90 ± 0.13 ^b	7.00 ± 0.66 ^{b,c,d,e}	6.72 ± 0.03 ^{e,f}
21	8.93 ± 0.14 ^a	7.19 ± 0.58 ^{b,c,d,e}	7.02 ± 0.60 ^{c,d,e}
Average	7.18 ± 1.22 ^A	6.25 ± 0.65 ^B	6.11 ± 1.35 ^B

The mean value followed by different letters showed a significant difference based on the Duncan's test at the 5% level (p -value < 0.05).

Table 1 shows that the garlic extract addition with different concentrations affects the increase in pH value of meat during storage. The statistical tests showed that the extract addition on active paper significantly affected the pH value ($p < 0.05$). Further test results showed that the treatment without the garlic extract (0%) was significantly different from the other treatments on average. However, the addition of 15% garlic extract was not significantly different from 20% garlic extract. Ponnampalam et al. stated that the pH of normal meat ranges from 5.5–5.7, and meat with a pH of >6 will be easily damaged by microorganisms, experiencing some deterioration such as color and texture aroma [44]. In Table 1, it can be seen that the initial pH value of fresh meat before storage was 5.68 and categorized as normal, and after storage it was classified as rotten on the 9th days of storage for treatment without garlic extract and on the 15th day for treatment with 15% and 20% garlic extracts.

3.2. Total Volatile Bases Nitrogen (TVBN)

The TVBN value is one of the factors used to measure the level of freshness in meat. Table 2 shows the changes in TVBN values in beef during cold storage.

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Table 2. The TVBN value of packaged meat samples stored in cold storage for 21 days.

Storage Time (Day)	Addition of Garlic Extract on Active Paper		
	0%	15%	20%
0	4.48 ± 0.00 ⁱ	4.48 ± 0.00 ⁱ	4.48 ± 0.00 ⁱ
3	4.76 ± 0.00 ⁱ	5.04 ± 1.40 ⁱ	4.58 ± 1.38 ⁱ
6	11.21 ± 1.46 ^s	4.20 ± 0.00 ⁱ	5.88 ± 0.56 ^{h,i}
9	27.27 ± 2.72 ^{d,e}	10.83 ± 0.65 ^s	9.43 ± 0.32 ^{g,h}
12	32.22 ± 9.54 ^c	22.32 ± 1.38 ^f	21.66 ± 2.83 ^f
15	34.18 ± 1.40 ^c	23.06 ± 0.71 ^f	22.97 ± 1.22 ^f
18	40.81 ± 1.17 ^b	24.65 ± 1.84 ^{e,f}	23.35 ± 1.71 ^{e,f}
21	55.28 ± 4.66 ^a	33.43 ± 1.13 ^c	30.26 ± 1.28 ^{c,d}
Average	26.27 ± 18.19 ^A	16.00 ± 11.26 ^B	15.33 ± 10.30 ^B

The mean value followed by different letters showed a significant difference based on the Duncan's test at the 5% level (p -value < 0.05).

Based on statistical tests, the addition of garlic extract on active paper significantly affected the TVBN value ($p < 0.05$). Further test results showed that the treatment without the extract (0%) addition was significantly different from the other treatments. However, 15% garlic extract was not significantly different from the 20% extract. TVBN criteria for identifying rotten beef products are >20 mg/100 g according to the Ministry of Agriculture and Forestry of Korea, 2016 [45]. The TVBN value of meat on the first day of storage was 4.48 mg N/100, indicating a fresh category (Table 2). On the 9th day of storage, meat treated without the extract had a TVBN value of 27.27 mg N/100 g, categorized as rotten. Meanwhile, the addition of 15% and 20% garlic extract with TVBN values of 22.32 mg N/100 g and 21.66 mg N/100 g had crossed the threshold on day 12. Based on the results obtained, the garlic extract affects the meat TVBN value during storage.

The increase in TVBN value is closely related to the decrease in beef quality. The treatment without garlic extract reached the threshold value earlier due to the higher microbial activity in the sample. The high number of microbes in the sample breaks down more meat protein to produce volatile ammonia compounds. This is consistent with Holman et al., where increased microbial value degrades protein rapidly during storage. Furthermore, it increases the amount of ammonia and other volatile compounds, as well as increases the TVBN value, where these compounds indicate the decay in meat [46]. The increase in the TVBN value of beef with treatment without and with the garlic extract was also different due to the allicin constituent and anti-microbial role. The inhibition of microbial growth minimizes the decomposition process of nutritional components in meat to reduce the TVBN value.

3.3. Total Bacteria of Total Plate Count (TPC) Method

The analysis determines the effect of using active paper on the total value of bacteria in meat during cold storage, as shown in Table 3.

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Table 3. The TPC value of packaged meat samples stored in cold storage for 21 days.

Storage Time (Day)	Addition of Garlic Extract on Active Paper		
	0%	15%	20%
0	3.52 ± 0.05 ⁱ	3.52 ± 0.05 ⁱ	3.52 ± 0.05 ⁱ
3	4.49 ± 0.04 ^{f,g,h}	4.16 ± 0.08 ^{g,h,i}	3.88 ± 0.06 ^{h,i}
6	6.49 ± 0.06 ^c	4.44 ± 0.03 ^{f,g,h}	4.29 ± 0.18 ^{g,h,i}
9	7.47 ± 0.02 ^b	5.08 ± 0.07 ^{e,f}	4.93 ± 0.48 ^{f,g}
12	8.30 ± 0.04 ^a	6.35 ± 0.71 ^{c,d}	6.09 ± 0.20 ^{c,d}
15	8.45 ± 1.22 ^{a,b}	6.43 ± 1.27 ^{c,d,e}	6.24 ± 0.69 ^{c,d}
18	8.17 ± 0.61 ^{a,b}	6.19 ± 0.15 ^{c,d}	5.69 ± 0.41 ^{d,e}
21	7.82 ± 0.16 ^{a,b}	5.81 ± 0.24 ^{c,d,e}	5.37 ± 0.54 ^{e,f}
Average	6.84 ± 1.87 ^A	5.25 ± 1.11 ^B	5.00 ± 1.02 ^B

The mean value followed by different letters showed a significant difference based on the Duncan's test at the 5% level (p -value < 0.05).

The garlic extract added to the active paper had a significant effect on the TPC value of meat ($p < 0.05$). Meanwhile, further test results showed that the treatment without garlic extract (0%) was significantly different from other treatments, but the addition of 15% was not significantly different from the 20% extract. The initial population of TPC meat was 3.52 log CFU/mL, which is categorized as fresh concerning microbiological quality (Table 3). However, this value increased to 6.49 log CFU/mL on the 6th day of storage in the control sample; this value is above the recommended maximum bacteriological limit, which was 6 log CFU/mL (SNI 3932:2008). The meat added with 15% and 20% garlic extract exceeded the maximum bacteriological limit on the 12th day of storage with TPC values of 6.35 log CFU/mL and 6.09 logs CFU/mL, respectively. Therefore, the addition of 15% and 20% extract as active packaging inhibits microbial growth and extends the shelf life of meat up to 6 days longer than the samples without the garlic extract in the active packaging. This is because garlic contains an anti-microbial compound in the form of allicin.

Garlic extract applied to the activated paper diffuses to the entire surface of the beef to inhibit the growth of bacteria through total and partial inhibition of RNA, DNA, and protein synthesis. Furthermore, allicin blocks bacterial enzymes with thiol groups to ultimately inhibit bacterial growth [47]. Mardiya also confirmed that the effectiveness of garlic extract with concentrations of 25%, 50%, and 100% was very effective in inhibiting *Staphylococcus aureus* bacteria and bactericidal [48]. Meanwhile, the extract with a concentration of 12.5% was declared as an inhibitor of bacterial growth (bacteriostatic), but less effective at inhibiting bacteria (bacteriocidal) [48].

The storage temperature of beef can also affect the meat TPC value. Meat cells in the postmortem phase experience metabolic reactions, which are highly dependent on storage temperature. According to Comi, the storage temperature and the metabolic reactions in meat are inversely proportional [49]. This is supported by Kuswandi and Nurfawaidi, where beef stored at room temperature is categorized as unfit for consumption at 10 h with a TPC value of 6.711 CFU/mL. Meanwhile, at cold temperature, it is categorized as unfit for consumption on day 7 with a TPC value of 6.961 CFU/mL [40].

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3.4. The Antibacterial Analysis Results of Active Paper

Antibacterial activity on active paper was tested against several bacteria that generally cause damage to beef, namely *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. Figure 2 illustrates the active paper's analytical findings for the bacterial inhibition zone region.

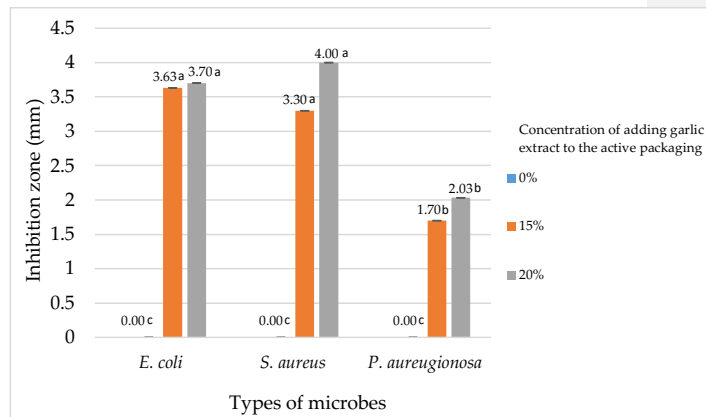


Figure 2. Diameter of bacterial growth barriers on activated paper. The mean value followed by different letters showed a significant difference based on the Tuckey's test at the 5% level (p -value < 0.05).

It can be seen from Figure 2 that with no addition of garlic extract to active paper (0%), the paper shows no inhibition zones to all three strains of bacteria tested. In comparison to active paper with the addition of both 15% and 20% garlic extract, it displayed bacterial inhibitory activity, implying that garlic extract possessed antibacterial activity.

According to Pan et al., the response to bacterial growth inhibition was divided into three categories: antibacterial activity with zones of inhibition greater than 6.00 mm (strong), 3–5 mm (moderate), and less than 3 mm (weak) [50]. Figure 2 provides information that shows active packaging with no addition of garlic extract had no inhibitory activity to all bacteria tested. While the addition of 15% and 20% of garlic extract to the active packaging showed a moderate response against *E. coli* (3.6 mm and 3.7 mm, respectively) and also moderate against *S. aureus* (3.7 mm and 4 mm, respectively). While for *P. aeruginosa* the active packaging categorized has a weak inhibitory response both in the administration of 15% and 20% garlic extract with 1.7 mm and 2 mm, respectively.

The diameter of the inhibitory zone created against *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* bacteria varies and could be due to the differences in bacterial structure that affect the penetration of allicin compounds through bacterial cell walls. The inhibitory zone formed by active packaging with the addition of both 15% and 20% garlic extract was classified as weak against *P. aeruginosa* due to the fact that gram-negative bacteria such as *P. aeruginosa* have complex cell walls, which prevent antibacterial compounds from penetrating the bacteria cell sufficiently to inhibit bacterial growth [51]. However, the findings of this study indicated that the application of garlic extract to the active package had the same inhibitory zone category against *E. coli* and *S. aureus*, even

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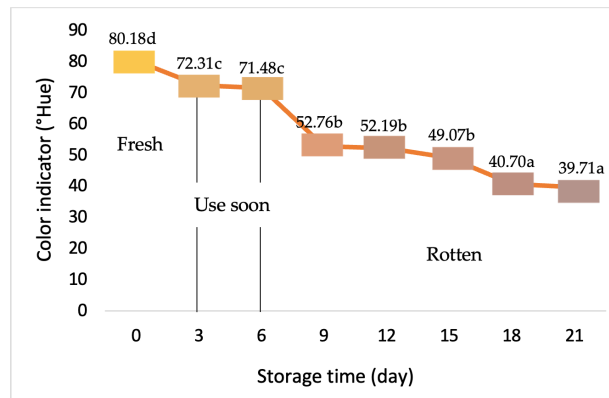
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though *E. coli* also includes Gram-negative bacteria. Even though the previous explanation for *P. aeruginosa* does not support the result obtained in this research for *E. coli*, these findings are consistent with the findings of Garba et al. and Safithri et al., who discovered that the inhibitory action to *E. coli* bacteria was typically more or equal to that of *S. aureus* [52-53].

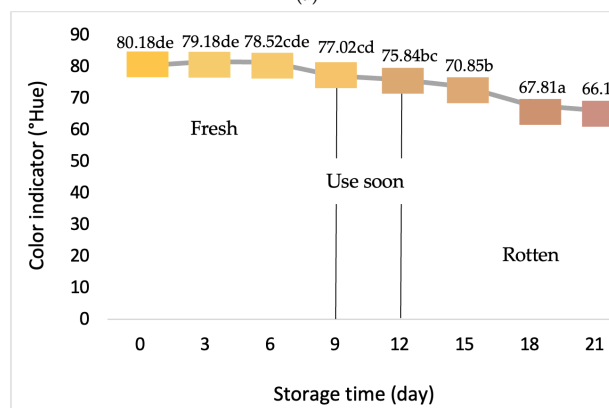
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3.5. Color Change Response on Intelligent Indicator Labels for Beef Packaged in Active Packaging at Cold Temperature Storage

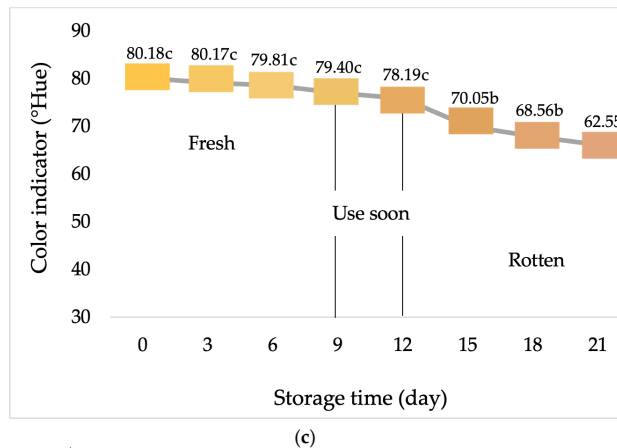
The color change analysis of the intelligent indicator determines the effect of adding garlic extract to active packaging with different concentrations on beef during storage at cold temperatures (chiller). The color change of the resulting indicator label is information of the freshness condition of the packaged beef. Significant color changes on indicators ease consumers to measure the freshness level based on visual appearance, as seen in Figure 3.



(a)



(b)



(c)

Figure 3. Color change profile of intelligent indicator labels with garlic extract addition on the concentrations at 0% (a), 15% (b), and 20% (c). The mean value followed by different letters showed a significant difference based on the Duncan's test at the 5% level (p -value < 0.05). Phase I with dark yellow indicating fresh, phase II with yellow and red gradations indicating have to be consumed soon, and phase III with faded red indicating rotten meat.

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Figure 3 showed that the °Hue value on the intelligent indicator label decreased during storage, from dark yellow to faded red. The color change is a response to the quality condition. There are three phases of change on intelligent packaging labels in the description.

The color change on the intelligent indicator label is closely related to the increased number of microbes and enzymes in meat during storage. These changes are caused by the decomposition of nutritional components of meat by enzymes and microbes, producing volatile base compounds used as an early sign of damage in packaged beef. This is consistent with Sani et al. who showed that volatile ammonia compounds in meat can increase the pH [54]. The increase in the pH of meat is caused by ammonia reacting with H^+ ions to produce H ions. The OH^- ions in the packaging are directly proportional to the pH value, affecting the change in intelligent indicator labels.

Delta E is a standard measurement for quantifying the difference between two colors. The standard delta E perception consists of five scales: 1.0 is invisible to the human eye; 1–2 visible through observation; 2–10 at a glance; 11–49 colors are more similar than opposite; and 100 colors are opposite [55]. Based on Figure 4, the smart indicator shows the value of delta E with a range of 1–18 for each treatment, except on storage days 3–6, the value of delta E is 0.77. This shows that smart indicators are generally easy to visualize based on color changes in smart packaging systems. Based on this, consumers will easily make visual assessments and also monitor the freshness of the meat through intelligent packaging sensors.

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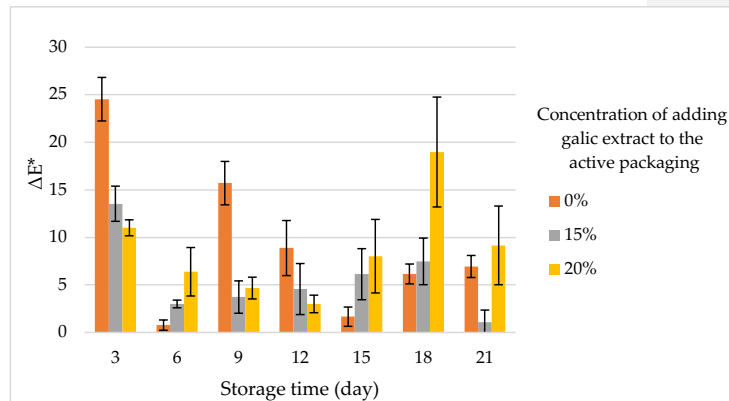
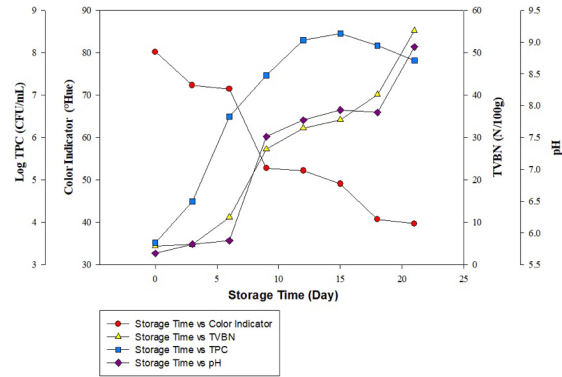


Figure 4. Total color difference (ΔE^*).

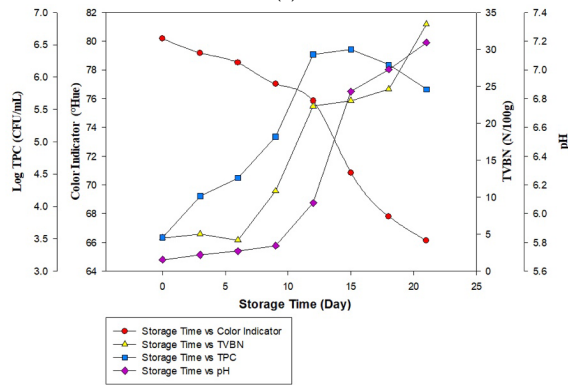
3.6. Correlation of Color Value Changes in Intelligent Packaging Indicators BTB: PR (1:1) with the Effect of Active Packaging on All Meat Spoilage Parameters

The relationship between the color change response of intelligent indicator labels with all test parameters on meat, such as TVBN, TPC, and pH, determined the sensitivity of intelligent indicator labels in detecting quality deterioration. This obtains synchronization between changes in the color of the intelligent indicator label to the deterioration of meat quality.

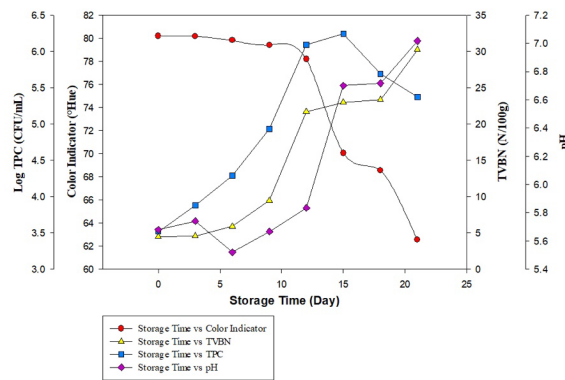
Based on Figure 5, beef stored without the garlic extract (0%) showed that the increase in the TVBN value and the pH of the meat was in line with the decrease in the color value of the intelligent packaging indicator. However, it is not consistent with the results of the TPC value since the beef TPC value crossed the threshold on the 6th day of storage, which is 6.4 CFU/mL. Meanwhile, the TVBN value is suitable for consumption on the 6th day. Mansur et al. also reported that the TVB-N values were inconsistent with the viable microbial count (TVC) [56]. Based on TVB-N, the samples were classified as damaged after 7 and 11 days of storage [56]. This is because the microbial calculation method is not selective against spoilage bacteria in meat but counts all the pathogenic microbes. Holman et al. also stated that the TVB-N value was more precise and accurate in classifying beef in fresh or rotten conditions than the total microbial value (TVC) [46].



(a)



(b)



(c)

Figure 5. Correlation of color change on intelligent indicator label with all beef spoilage parameters with the garlic extract addition: (a) control, (b) 15%, and (c) 20%.

Based on Figure 5b,c, beef stored with 15% and 20% garlic extract treatment showed that the increase in the parameters of the meat quality deterioration, including TPC, TVBN, and pH, was consistent with the decrease in the color of the intelligent indicator label. The intelligent indicator changes color from yellow (fresh) to faint red (rotten) during storage. The increase in TVBN, TPC, and pH values is influenced by the high nutritional content, specifically protein broken down into polypeptides and other amino acids through the deamination process to form ammonia. With the increase in microbes, the nutritional components decomposed by microbes are also greater and produce basic volatile compounds, increasing the pH value of meat during storage. The resulting volatile compounds accumulate in the packaging and are directly detected by the intelligent indicator label. The reduction in the color value of the intelligent indicator label shows the presence of many alkaline volatile chemicals in the package and the rottenness and unfitness for the ingestion of the meat. Furthermore, when viewed for the visual appearance on the 6th and 12th day of storage without (0%) and with (15% and 20%) garlic extract, the beef was damaged and unfit for consumption, which is characterized by the formation of mucus, emitting a sour/rotten smell, and the texture beginning to be sticky to the touch.

The increase in the beef quality deterioration parameter value for each treatment was different due to the addition of garlic extract as active packaging. The extract has a bioactive compound in allicin to inhibit microbial growth. This compound in the package diffuses in the meat surface to suppress the microbe growth. According to Deresse, it works by completely inhibiting bacterial RNA synthesis and inhibiting DNA and protein synthesis [47]. The pattern of relationship between the parameters of the decline in beef quality and the color analysis value of indicator labels is also consistent with Kuswandi and Nurfawaidi, where a linear relationship pattern to the parameters of the beef freshness level with changes in the color intensity of the bromocresol purple and methyl red intelligent labels was observed [40]. In this study, beef treated without the garlic extract experienced a decrease in quality and was not suitable for consumption on the 6th day. Meanwhile, with 15% and 20% garlic extract, the meat was damaged and unfit for consumption on the 12th day of storage. Previous research applying garlic extract as active packaging in meat was performed and resulted as the 15% garlic extract enhancing the shelf life of meat to 16 h from 12 h without garlic extract at room temperature [57]. This indicates that garlic extract storing at cold temperatures can extend the shelf life of packaged meat. Visually, the intelligent indicator label used Whatman paper no. 1 as the base material, and the BTB + PR (pH 5.00) used in intelligent packaging for beef freshness, with a color change from yellow (beginning) to faint red (late). Therefore, it assists customers in determining freshness without touching the texture or opening the packaging.

4. Conclusions

The garlic extract in active packaging, supported by the microbiological evaluation results of TVBN and pH, extended shelf life during storage. In addition, the results showed that the active paper with 15% and 20% garlic extract extended the shelf life to 12 days of cold storage. The color change profile of the intelligent indicator label combination of BTB + PR pH 5.00 was easily observed from dark (fresh) to reddish yellow (to be consumed immediately). The red color fades

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when the beef is rotten and unfit for consumption. Therefore, the combination of active and intelligent packaging is suggested as a potential packaging application in fresh meat packaging.

Author Contributions: Conceptualization, A.D.; methodology, A.D. and A.F.A.; software, A.F.A.; validation, A.D. and M.D.; formal analysis, A.D. and A.F.A.; investigation, A.F.A.; resources, A.D.; data curation, A.D. and A.F.A.; writing—original draft preparation, M.D. and A.F.A.; writing—review and editing, A.D., M.D. and A.F.A.; visualization, A.F.A.; supervision, A.D.; project administration, A.D. All authors have read and agreed to the published version of the manuscript.

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Commented [M33]: Newly added information. Same as below, Please check all highlights and confirm.

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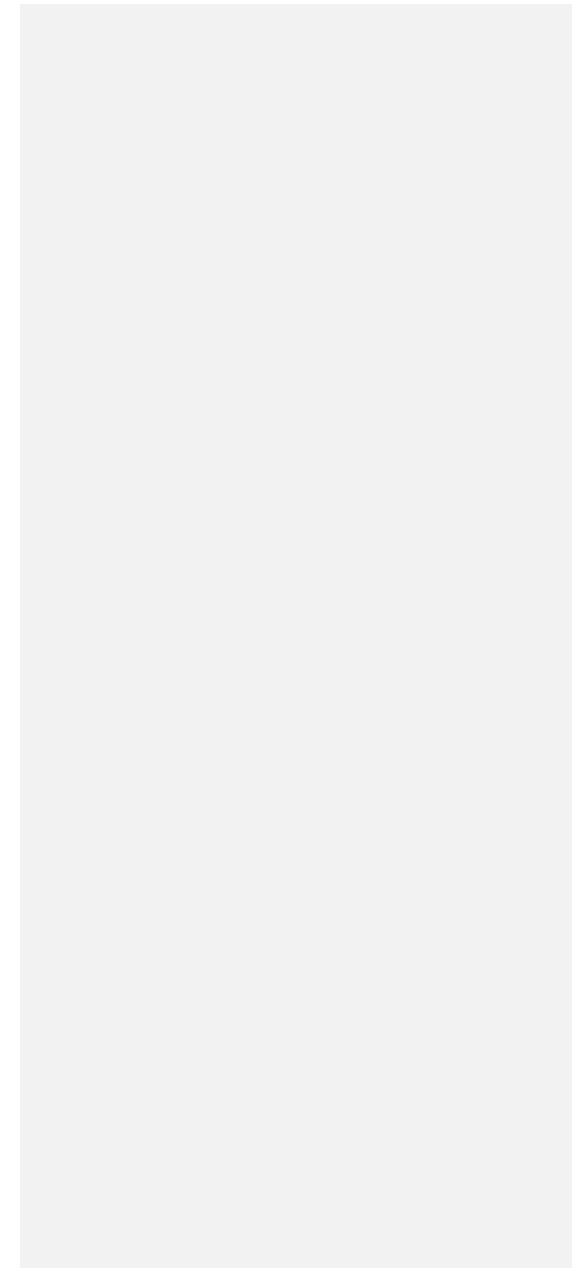
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Type of manuscript: Article

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Authors: Andi Dirpan *, Muspirah Djalal, Andi Fadiah Ainani

Received: 30 April 2022

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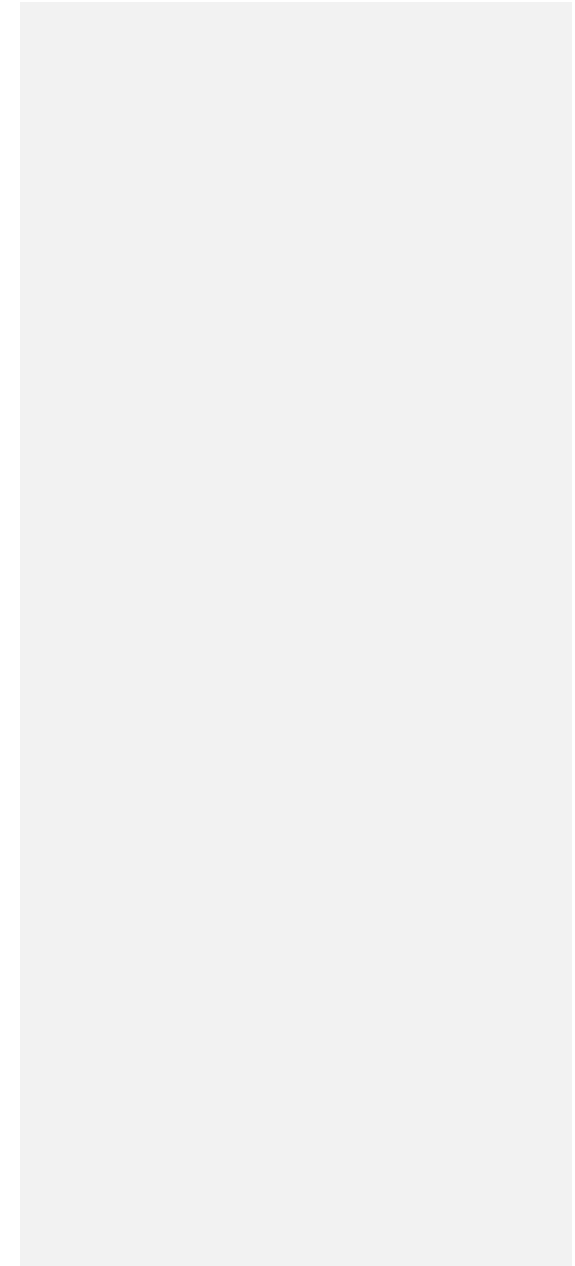
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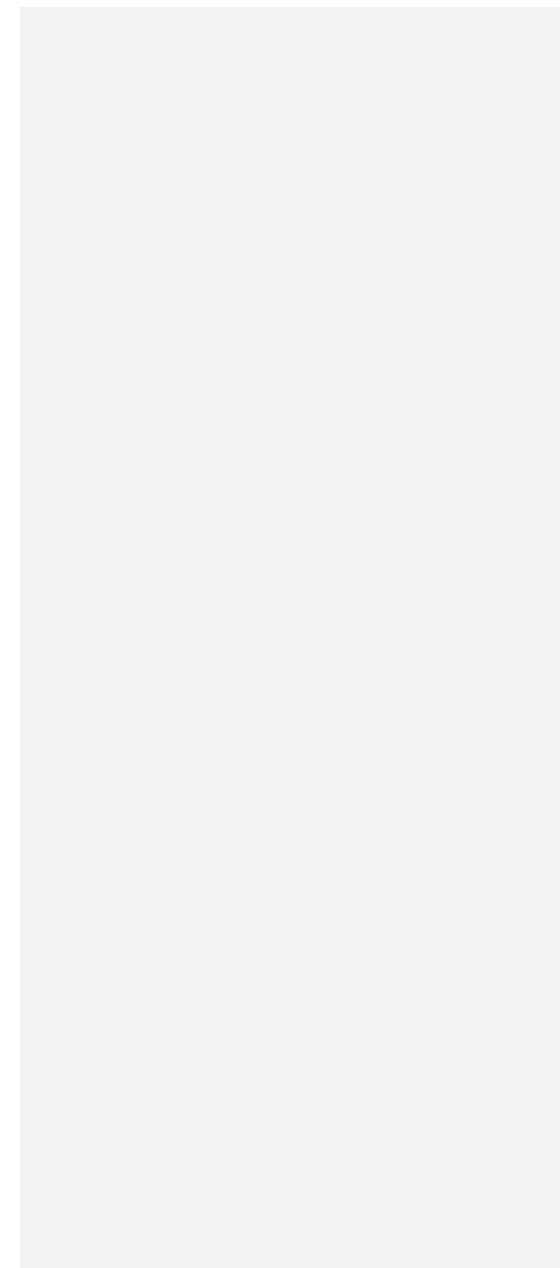
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